MISCELLANEOUS NOTES

7. HOST PREFERENCE AND REPORT OF TWO NEW HOST PLANTS OF LORANTHUS LONGIFLORUS AT INDORE, MADHYA PRADESH, INDIA

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Introduction

Species of the Mistletoe family characteristically show parasitism. Loranthus is recorded from Sumatra, New Guinea, Ceylon (now Sri Lanka), Pakistan (Abdulla 1973), India, Thailand, Australia, China, Bangladesh, Malaysia, Myanmar (Pattanayak et al. 2008) and United Provinces (India) (Srivastava 1935). Loranthus longiflorus is distributed throughout India (Chopra 1975). A large number of host plants of L. longiflorus are reported by various scientists (Srivastava 1935; Sampathkumar and Kunchithapatham 1969; Narayanasamy and Sampathkumar 1981; Sampathkumar and Selvaraj 1981; Balsubramanian and Sugathan 1986; Sawant et al. 2008).

Study Area and Method

Indore is situated at 22° 48' N and 75°48' E, 553 m above mean sea level. The campus of Government Holkar Science College (an area of about 36 acres) and its adjoining area and Vasudeonagar (an area about 5 acres). Indore, Madhya Pradesh, were selected to survey the host range of *Loranthus longiflorus*. 762 flowering plants of 60 species were screened between January 15 and April 15, 2009. This is a common species in the urban area of Indore.

Results and Discussion

Loranthus longiflorus Desr. flourished on 7 out of 43 species in the study area. Two host plants: Albizia lebbeck

Table 1: Host range relationship of Loranthus longiflorus in the campus and adjoining area of Government Holkar Science College, Indore, M.P.

S. No.	Plant Studied	Family	Total no. of studied	Total no. of infected	S. No.	Plant Studied	Family	Total no. of studied	Total no. of infected
1.	Aegle marmelos	Rutaceae	2	0	23.	Pithecellobium	Mimosaceae	47	1
2.	Acacia nilotica	Mimosaceae	57	0		dulce			
3.	Albizia lebbeck	Mimosaceae	16	10	24.	Pterospermum	Sterculiaceae	1	0
4.	Annona squamosa	Annonaceae	4	0		acerifolium			
5.	Azadirachta indica	Meliaceae	60	5	25.	Phoenix sylvestris	Palmae	13	0
6.	Artocarpus	Moraceae	1	0	26.	Mitragyna parviflora	Rubiaceae	4	0
	heterophyllus				27.	Santalum album	Santalaceae	68	0
7.	Butea monosperma	Papilionaceae	1	0	28.	Polyalthia longifolia	Annonaceae	20	0
8.	Ceiba pentandra	Bombacaceae	9	0	29.	Tamarindus indica	Caesalpiniaceae	23	0
9.	Caesalpinia	Caesalpiniaceae	8	0	30.	Ziziphus mauritiana	Rhamnaceae	10	0
	pulcherrima				31.	Kigelia pinnata	Bignoniaceae	1	0
10.	Cordia myxa	Boraginaceae	16	2	32.	Dalbergia sissoo	Papilionaceae	2	0
11.	Cassia fistula	Caesalpiniaceae	11	0	33.	Madhuca	Sapotaceae	2	0
12.	Delonix regia	Caesalpiniaceae	13	0		longifolia			
13.	Emblica officinalis	Euphorbiaceae	9	0	34.	Pongamia pinnata	Papilionaceae	12	0
14.	Eucalyptus	Myrtaceae	46	1	35.	Tectona grandis	Verbenaceae	1	0
	camaldulensis				36.	Bombax	Bombacaceae	7	0
15.	Syzygium cumini	Myrtaceae	41	0		malabaricum			
16.	Ficus benghalensis	Moraceae	3	0	37.	Bauhinia variegata	Caesalpiniaceae	1	0
17.	Ficus racemosa	Moraceae	7	0	38.	Grevillea robusta	Proteaceae	9	0
18.	Ficus religiosa	Moraceae	18	1	39.	Millingtonia	Bignoniaceae	3	0
19.	Leucaena	Mimosaceae	29	0		hortensis			
	leucocephala				40.	Acacia	Mimosaceae	1	0
20.	Mangifera indica	Anacardiaceae	40	15		leucophloea			
21.	Mimusops elengi	Sapotaceae	6	0	41.	Morinda tinctoria	Rubiaceae	3	0
22.	Peltophorum	Caesalpiniaceae	46	0	42.	Cassia siamea	Caesalpiniaceae	7	0
	acerifolium				43.	Ixora parviflora	Rubiaceae	3	0

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(62.5%) and Mangifera indica (37.5%) seem to be most susceptible (Table 1). However, in Vasudeonagar colony 10 out of 29 species were affected. Here, Mangifera indica (58,33%) and Grevillea robusta (44,44%) were the preferred hosts of Loranthus. Two new hosts, i.e., Pseuderanthemum atropurpureum (Acanthaceae) and Lagerstroemia indica (Lythraceae) (Table 2), were observed during this study. Occurrence of Loranthus on these shrubs, grown as ornamentals, proves that it has tremendous adaptability for hosts. L. longiflorus generally prefers big trees with welldeveloped lenticels on bark, but in this case the condition did not appear to be applicable. Infection of Loranthus on these shrubs appeared to be by chance. Another important feature recorded during the study was that L. longiflorus does not grow on monocots, probably as monocots have scattered vascular bundles and penetration of Loranthus roots in the xylem of the host may be difficult.

Pseuderanthemum atropurpureum is nearly 3.65 m tall having several (38 to 40) thick and thin branches arising from the stem at ground level. Loranthus had attacked a 0.9 cm diameter branch. Interestingly, the apical portion of this branch was dead after infection. This plant was c. 12.19 m away from three infected plants. Another host Lagerstroemia indica, was c. 3.05 m tall. c. 6.09 m from two infected plants. More than 300 species of host plants of Loranthus longiflorus have been reported in literature (Srivastava 1935; Suryaprakash et al. 1967; Sampathkumar and Kunchithapatham 1969; Narayanasamy and Sampathkumar 1981; Sampathkumar and Selvaraj 1981; Indrani and Balasubramanian 1985; Balsubramanian and Sugathan 1986; Ramchandranair and Krishanakumar 1989; Pattanayak et al. 2008; Sawant et al. 2008), but none have reported these two hosts. Therefore, these two species are new reports for India. In our view, this parasite is not host specific; infection success depends on favourable conditions at the time the seed falls on the host. Narayanasamy and Sampathkumar (1981) report higher osmotic pressure of semi-parasite than hosts for infection success

Loranthus longiflorus is one of the most important traditional medicinal species. Its medicinal properties are greatly influenced by the host plant (Jain 1997; Pattanayak *et al.* 2008), therefore, further study is required for evaluation of *Loranthus longiflorus* on the new reports.

Table 2: Host r	ange relationship of Loranthus longiflorus	
in	Vasudeonagar, Indore, M.P.	

S. No.	Plant Studied	Family	Total no. of studied	Total no. of infected plants
1.	Cassia fistula	Caesalpiniaceae	3	1
2.	Lagerstroemia indica	Lythraceae	2	1
3.	Plumeria alba	Apocynaceae	3	0
4.	Peltophorum acerifolium	Caesalpiniaceae	3	0
5.	Pongarnia pinnata	Papilionaceae	2	0
6.	Azadirachta indica	Meliaceae	3	1
7.	Delonix regia	Caesalpiniaceae	1	0
8.	Millingtonia hortensis	Bignoniaceae	3	0
9.	Grevillea robusta	Proteaceae	9	4
10.	Moringa oleifera	Moringaceae	2	0
11.	Psidium guajava	Myrtaceae	7	2
12.	Mangifera indica	Anacardiaceae	12	7
13.	Ficus religiosa	Moraceae	1	0
14.	Annona reticulata	Annonaceae	1	1
15.	Michelia champaca	Magnoliaceae	1	0
16.	Bambusa sp.	Bombacaceae	1	0
17.	Cocos nucifera	Palmae	2	0
18.	Gardenia jasminoides	Rubiaceae	1	0
19.	Cassia renigera	Caesalpiniaceae	1	0
20.	Cordia myxa	Boraginaceae	1	0
21.	Hibiscus rosa-sinensis	Malvaceae	2	0
22.	Ficus racemosa	Moraceae	1	1
23.	Terminalia catappa	Combretaceae	2	0
24.	Bauhinia variegata	Caesalpiniaceae	1	0
25.	Callistemon lanceolatus	Myrtaceae	1	1
26.	Alstonia scholaris	Apocynaceae	11	0
27.	Melia azadirachta	Meliaceae	2	0
28.	Pseuderanthemum atropurpureum	Acanthaceae	1	1
29.	Thevetia neriifolia	Apocynaceae	1	0

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8. SELAGINELLA RETICULATA (HOOK. & GREV.) SPRING (SELAGINELLACEAE) – A NEW RECORD TO THE PTERIDOPHYTIC FLORA OF RAJASTHAN, NORTH-WESTERN INDIA

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Rajasthan in north-west India is the largest state with an area of 3,42,274 sq. km and lies between $23^{\circ}3'-30^{\circ}12'$ N and $69^{\circ}30'-78^{\circ}17'$ E. Aravalli ranges, which are one of the oldest mountain ranges of the world, diagonally divide the state into two distinct climatic regions. The region towards the north-western side of Aravalli is a desert or semi-desert characterized by sand dunes, high wind velocity, high temperature and thorny vegetation. The region towards the south-eastern side is a humid zone with hills of variable heights, ravines, plains, rivers and dense forests.

Sitamata Wildlife Sanctuary (Fig. 1) is one of the protected areas of Rajasthan with a wide range of habitats in the Aravalli ranges. It is exceptional for diversity and interspersed habitats, which includes areas of teak stands, perennial streams, sloping hills and fine groves of mixed woodlands. The Sanctuary lies on the Udaipur-Pratagarh state highway at a distance of 100 km from Udaipur and 40 km from Prataggarh. The major part of the Sanctuary lies in the newly formed civil district of Prataggarh, only 74.21 sq. km comes under district Chittorgarh. The Sanctuary covers an area of 422.95 sq. km (195.09 sq. km core area and 227.86 sq. km buffer area) and lies between 24° 04'-24° 23' N and 74° 25-74° 40' E. The occurrence of flying squirrel is the greatest attraction of the Sanctuary.

Geographically, there are three major operating systems, namely the Aravalli, the Vindhyan and Malva plateau, which result in a variety of habitats. The annual rainfall in the Sanctuary area ranges from 664.60 mm to 1,430.20 mm with an average 30 rainy days. The temperature varies from 18.6-33.6 °C.

Selaginella Beauv, with 700 species (Pichi-Sermolli 1977) is represented by 62 species in India (Alston 1945; Panigrahi and Dixit 1966, 1967, 1968; Dixit 1984, 1992). The first enumeration of the Indian species of Selaginella was provided by Alston (1945). Subsequently, Panigrahi and Dixit (1966, 1967, 1968) carried out prolonged research on this genus and dealt with 30 species of Selaginella in detail. Sharma and Bhardwaja (1976) have reported the occurrence of Selaginella repanda (Desv.) Spring for the first time from Gwaparnath, Kota (south-east Rajasthan). A new species of Selaginella, namely Selaginella rajasthanensis was described by Gena et al. (1979) from Kunda Khoh. Shahabad, Baran district of Rajasthan. Recently, Dulawat and Chaudhary (2008) have recorded Selaginella ciliaris (Retz.) Spring from Sitamata, Rajasthan. The present paper deals with the detailed taxonomic account, distribution and ecological notes of Selaginella reticulata recorded from Sitamata forest in Rajasthan.

An extensive survey of the area of Sitamata Wildlife Sanctuary was carried out during 2004-2009. Various localities which seem likely to support the growth of *Selaginella* were visited regularly, especially during rainy season (July-October). Field observations, such as habit, habitat, associated plants were recorded. Identification was done using Alston (1945), Panigrahi and Dixit (1966), and Dixit (1984, 1992). Morphological observations and camera